

SCIENTIFIC REPORTS

OPEN

Rediscovery and reclassification of the dipteran taxon *Nothomicrodon* Wheeler, an exclusive endoparasitoid of gyne ant larvae

Received: 07 November 2016

Accepted: 28 February 2017

Published: 31 March 2017

Gabriela Pérez-Lachaud¹, Benoit J. B. Jahyny^{2,3}, Gunilla Ståhls⁴, Graham Rotheray⁵, Jacques H. C. Delabie⁶ & Jean-Paul Lachaud^{1,7}

The myrmecophile larva of the dipteran taxon *Nothomicrodon* Wheeler is rediscovered, almost a century after its original description and unique report. The systematic position of this dipteran has remained enigmatic due to the absence of reared imagos to confirm identity. We also failed to rear imagos, but we scrutinized entire nests of the Brazilian arboreal dolichoderine ant *Azteca chartifex* which, combined with morphological and molecular studies, enabled us to establish beyond doubt that *Nothomicrodon* belongs to the Phoridae (Insecta: Diptera), not the Syrphidae where it was first placed, and that the species we studied is an endoparasitoid of the larvae of *A. chartifex*, exclusively attacking sexual female (gyne) larvae. *Nothomicrodon* parasitism can exert high fitness costs to a host colony. Our discovery adds one more case to the growing number of phorid taxa known to parasitize ant larvae and suggests that many others remain to be discovered. Our findings and literature review confirm that the Phoridae is the only taxon known that parasitizes both adults and the immature stages of different castes of ants, thus threatening ants on all fronts.

Ants are hosts to at least 17 orders of myrmecophilous arthropods (organisms dependent on ants), ranging from general scavengers to highly selective predators and parasitoids that attack either ants, their brood or other myrmecophiles^{1–3}. Recent reviews of ant and myrmecophile relationships reveal both diversity and complexity^{4–10}. The communities inside ant nests and colonies have been likened to homeostatic fortresses or microcosms that encapsulate phenomena normally encountered only at larger scales¹¹. Least studied, however, are ant parasitoid relationships. Compared to other myrmecophiles, few ant parasitoids appear to be entirely successful in evading host colony defense mechanisms^{12,13}. Unlike other myrmecophiles, ant parasitoids do not integrate with the host colony and have to deal with issues such as locating and successfully parasitizing suitable hosts, and later escaping from the host nest. Some ant parasitoids have mechanisms that are rare in other parasitoids, such as oviposition away from the host combined with a freely mobile, first instar larva (*planidium*) that completes the initial stage of parasitization^{12,14}. Ant parasitoids can also manipulate host behavior such as provoking in-fighting between worker ants through semiochemicals released by ovipositing females¹⁵, inducing nest leaving in parasitized workers by developing parasitoids¹⁶, or reducing host aggressiveness by emerging imagos¹⁷.

About 750 species, from five taxa of Ecdysozoa (four in Arthropoda and one in Nematoda), are ant parasitoids¹⁸. Most belong to the Hymenoptera, and a diverse array of families are involved¹². Dipterans also parasitize ants and representatives of four families are primary parasitoids. Most belong to the Phoridae, for example the

¹El Colegio de la Frontera Sur, Av. Centenario Km 5.5, Chetumal 77014, Quintana Roo, Mexico. ²Universidade Federal do Vale do São Francisco UNIVASF, Colegiado de Ciências Biológicas, Campus Ciências Agrárias - Rodovia BR 407, 12 Lote 543 Petrolina, Pernambuco, Brazil. ³Seção de Entomologia, Comissão Executiva do Plano da Lavoura Cacaueira, Centro de Pesquisa do Cacau (CEPLAC, CEPEC), Cx.P.7, 45600-970, Ilhéus, Bahia, Brazil. ⁴Finnish Museum of Natural History, Entomology Dept., P.O. Box 17, FIN-00014 University of Helsinki, Finland. ⁵National Museums Scotland, West Granton Road, Edinburgh, EH5 1JA, United Kingdom. ⁶Laboratório de Mirmecologia, Convênio CEPLAC/UESC, Cocoa Research Center (CEPEC), 45600-000, Itabuna, Bahia, Brazil. ⁷Centre de Recherches sur la Cognition Animale, Centre de Biologie Intégrative, Université de Toulouse UPS, CNRS-UMR 5169, 118 Route de Narbonne, 31062 Toulouse Cedex 09, France. Correspondence and requests for materials should be addressed to J.-P.L. (email: jean-paul.lachaud@univ-tlse3.fr)

so-called ant-decapitating flies of the genera *Apocephalus*, *Pseudacteon* or *Neodohrniphora* that mostly parasitize workers¹⁹. Single parasitic species also exist in the Tachinidae (*Strongygaster globula*, an endoparasitoid of colony-founding queens of the genus *Lasius* (Formicinae)²⁰), the Syrphidae (*Hypselosyrphus trigonus*, an ectoparasitoid of prepupae of the arboreal ant *Neoponera villosa* (Ponerinae)²¹), and the Chloropidae (*Pseudogaurax paratolmos*, an ectoparasitoid of the larvae of the fungus-growing ant *Apterostigma dentigerum* (Myrmicinae)²²).

Almost a century ago the enigmatic taxon *Nothomicrodon aztecarum* (Wheeler, 1924) was raised on the basis of morphologically unusual larvae found among the brood of a carton nest ant, *Azteca trigona* (Dolichoderinae), from Barro Colorado Island, Panama²³. No adult *N. aztecarum* were obtained and due to the similarity in larval stages, Wheeler placed *Nothomicrodon* in the Syrphidae as an ally of the genus *Microdon* whose larvae are well known predators of ant brood²⁴. The true affinity of *Nothomicrodon* has remained unresolved because adults have not been reared and larvae have not been re-encountered since its original description. Historically, the genus has been treated as *incertae sedis* by syrphid experts. Cheng & Thompson²⁵ stated that the larva has none of the characteristics of microdontine larvae (flattened creeping sole, convex dorsal surface) and based on a suggestion from G.E. Rotheray, speculated that it might belong to the Phoridae. The most up-to-date revision of Microdontinae also treated *Nothomicrodon* as an unplaced taxon²⁶.

In this paper we report on the rediscovery of *Nothomicrodon*. Scrutiny of entire nests of *Azteca chartifex* collected in Brazil combined with morphological and molecular studies enabled us to establish beyond doubt that these larvae belong to the Phoridae and that they were endoparasitoids of *A. chartifex* larvae, more specifically of the sexual female (gyne) larvae. Based on these data and a literature review of phorid parasitoids attacking social insect brood, we confirm that the Phoridae is the only insect family known with species that parasitize both the adults and immature stages of their ant hosts, thereby threatening ants on all fronts.

Results

DNA sequencing and identification. The obtained COIa fragment comprised 653 bp and the COIb 780 bp. The top 20 closest matches of the COIa sequence identification on BOLD were all Phoridae samples (except one Agromyzidae (Diptera: Opomyzoidea) sample). The highest similarity, 88.7%, was found with an unpublished Phoridae sample. Similarities of 88.5% were found with published barcodes of two phorid flies from USA and Canada (BIN BOLD: AAM9347 and BOLD: AAN8679), both from the genus *Megaselia*. Blasting the COIb fragment in GenBank (www.ncbi.nih.gov, on 7 March, 2016) returned a long list of Phoridae samples as closest matches. Sequence similarity of 84% was reported for a sample of the phorids *Anevrina variabilis* (GenBank accession number GU559934) and *Dohrniphora cornuta* (HM352592). Sequence similarity of 83% was reported for two samples of the phorid *Apocephalus paraponerae* (AF217478-9) which is a known ant parasitoid²⁷. No syrphid fly species appeared in the top 20 closest matches for both COIa and COIb sequence identification. Moreover, the neighbor-joining and maximum likelihood analyses placed the *Nothomicrodon* sequence among all included samples of other Phoridae (Fig. 1, Table S1), herewith confirming the identification of the sample as a phorid fly, not a syrphid.

Description of *Nothomicrodon* third instar larva (n = 2). *Overall appearance.* Pear-shaped with a broad, oval-shaped abdomen and a narrow, tapering thorax; pale to brown with a coriaceous integument (Fig. 2A); abdomen smooth except for a single pair of deep infolds across the dorsum (see Fig. S1A); head skeleton with the apex of the labium external to the fleshy pseudocephalon and comprising a pair of downwardly projecting, black, heavily sclerotized hooks, rest of the head skeleton translucent (see Fig. S2A), poorly sclerotized and lacking cibarial ridges.

Description. Length about 4.5 mm (1.5 mm pseudocephalon and thorax + 3 mm abdomen), abdomen 3.5 mm wide and maximum height about 0.75 mm; width of the metathorax where it joins the broader abdomen, about a quarter the maximum width of the abdomen and at the prothoracic apex, about a fifth the width of the base of the metathorax (see Fig. S1B,C); antennae on the dorso-lateral margins of the pseudocephalon, just posterior to the apex, appearing as a pair of cylindrical, tapering structures (see Fig. S2B), maxillary palpi not recognizable in the specimens examined; ventrally, apex of pseudocephalon with a pair of inwardly directed, flange-like, cuticular projections (see Fig. S1D); pseudocephalon and thorax retractile, as revealed by folds and creases along which the integument probably collapses and/or retracts, by analogy with other larvae²⁸; probable margin between the pseudocephalon and prothorax indicated by a deep infold; prothorax elongate, about twice as long as each of the pseudocephalon, mesothorax and metathorax which are all of a similar length (see Fig. S1C); towards the rear of the prothorax on the dorso-lateral margins, are the anterior respiratory processes comprising a pair of cylindrical projections inclined forward and with the spiracles across the apex (see Fig. S1C); metathorax attached to a firm infold at the apex of the first abdominal segment by a band of thin, flexible integument; lateral and posterior margins of the abdomen pinched and with a slight, continuous beading; externally segments marked only by segmental pattern of inconspicuous sensilla, each accompanied by a single hair-like seta, abdomen otherwise unmarked except for the third abdominal segment whose boundaries with adjacent segments are marked by deep infolds across the dorsum (see Fig. S1A); anal segment crescent shaped, as revealed by the pattern of sensilla round the slight, dome-shaped posterior respiratory process; this process with four pairs of short, parallel spiracles orientated dorso-ventrally, above which are a pair of cuticular scars, the paired spiracular plates separated mid-apically by a longitudinal, slit-like infold (see Fig. S2C); entire body coriaceous, locomotory organs apparently absent; head skeleton (see Fig. S2A) 0.5 mm long, form typical for a member of the Platypezoidea²⁸; sclerotization slight except for the black, sclerotized apex to the ventral, labial arm which projects externally from the apex of the fleshy pseudocephalon in the form of a pair of stout, downwardly projecting hooks; apex of labrum and mandibles tapered, inconspicuous and insignificant relative to the much larger labial hooks; ventral and dorsal cornua elongate and parallel, not diverging; ventral cornu slightly broader than dorsal cornu; cibarial ridges absent.

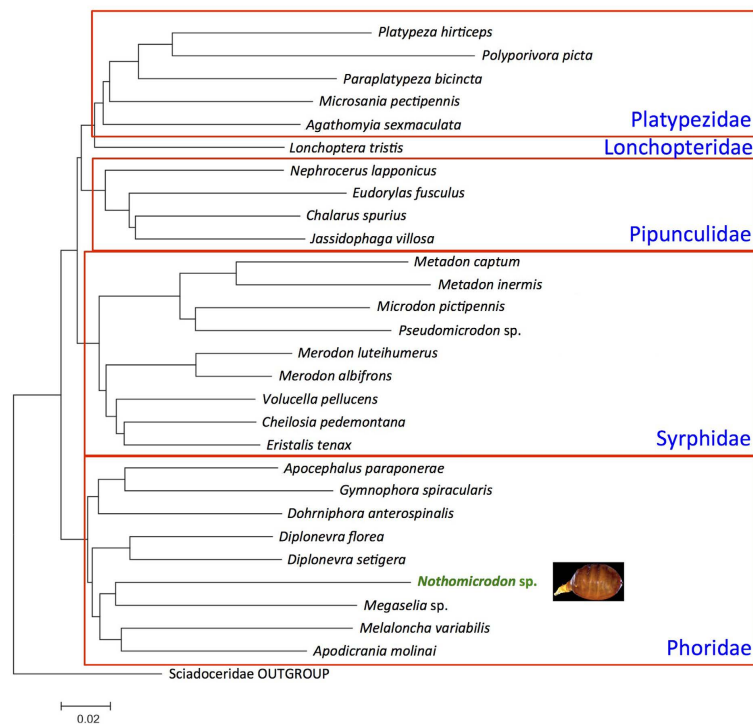


Figure 1. Results of the Neighbor-Joining analysis based on mtDNA COIb sequences. Photo: H. Bahena Basave.

Taxonomic notes. The third instar larva of our species agrees well with the description and dimensions of the larva of *N. aztecarum*²³, and we consider it congeneric with this species.

Life cycle and developmental stages of *Nothomicrodon*. Parasitized *A. chartifex* larvae, in both early and advanced stages of parasitoid development, can be identified by the small, oval, melanized/sclerotized scar from which the posterior respiratory process of the parasitoid projects from the host cuticle (Fig. 3A). Advanced stages of development (third instar *Nothomicrodon* larvae) are easily observed through the host integument (Fig. 2B,C). Breathing holes may be located on any part of the ant larva including the head. The hole is round-oval and measures 0.07 mm in diameter on average ($n = 7$); its rim is raised and heavily sclerotized. Upon host dissection, eggs were found firmly attached to the host cuticle (Fig. 3B,C, $n = 2$ cases, Table 1). Eggs are elliptical in form with the apical portion more acute than the base. One egg was measured: length = 1.0 mm, base = 0.37 mm and apical portion = 0.19 mm. All developmental stages of *Nothomicrodon* remained attached by the posterior respiratory process to the host cuticle. As with other phorid species²⁹, *Nothomicrodon* larva has three instars. The first and second are of a whitish color and the cuticle is not sclerotized (Figs 3D and 4A). First/second instar *Nothomicrodon* larvae dissected from the host measured 1.34 ± 0.09 mm (mean \pm SD) in width and 1.88 ± 0.11 mm in length ($n = 4$). Three of these larvae had the pseudocephalic region extended, length 0.56 ± 0.12 mm. Early third instar larvae are yellow in color (Fig. 4B) and the cuticle has already the leathery aspect of the fully grown, reddish-dark brown third instars (see Fig. 2A). After feeding is completed, third instar larvae cut open the host cuticle with their labial hooks (Fig. 4C). These larvae measure 3.02 ± 0.25 mm in width and 3.51 ± 0.14 mm in length (mean \pm SD; $n = 8$).

Host caste and developmental stage targeted. *Azteca chartifex* larvae are oval in form and practically hairless; the mouthparts are small, the mandibles are feebly sclerotized and, as in other dolichoderine taxa, mobility is almost lost³⁰. Gyne larvae of the Dolichoderinae subfamily are much larger than worker larvae³⁰. The length and width of a random sample of larvae of varying sizes were obtained ($n = 212$). The MDA model discriminated parasitized from unparasitized larvae according to their attributes, with parasitized larvae exclusively in the larger size class, which corresponds to gyne larvae (Fig. 5, see Fig. S3). The model explained 89 and 100% of the between group variance of the variables, and correctly assigned most of the larvae (deviance 19.8, misclassification error 0.94%). Only one parasitized and one unparasitized larvae were not correctly assigned. Parasitized larvae measured on average 3.4 ± 0.3 mm in width and 4.7 ± 0.5 mm in length (mean \pm SD; $n = 25$); unparasitized larvae measured on average 1.5 ± 0.4 mm in width and 2.1 ± 0.6 mm in length ($n = 187$). *Nothomicrodon* was found only in the nests that contained gyne larvae: no small or fully-grown minor or major worker larvae or male larvae were parasitized.

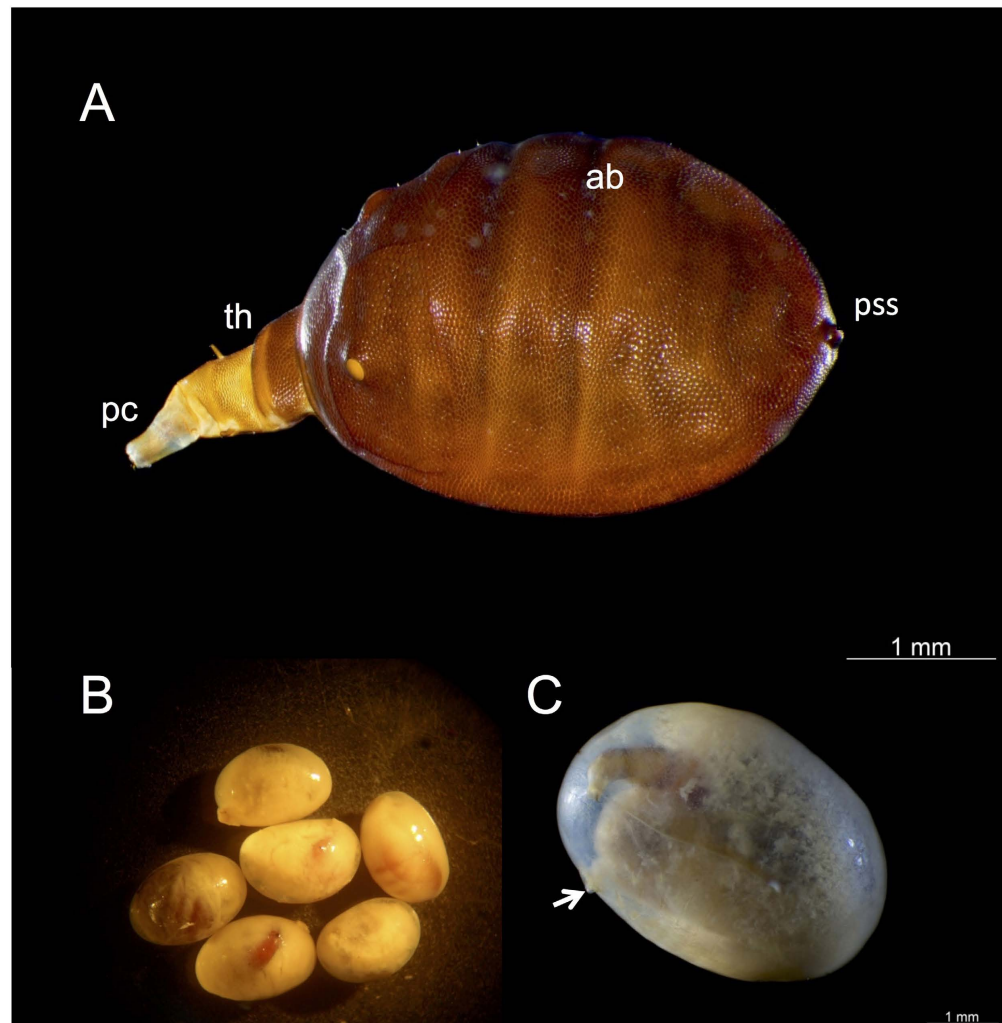


Figure 2. *Nothomicrodon* third instar larva. (A) General aspect of whole larva; ab: abdomen; pc: pseudocephalon; pss: posterior spiracular system; th: thorax. (B) General aspect of *A. chartifex* gyne larvae parasitized by *Nothomicrodon* (C) Fully grown *Nothomicrodon* larva inside an *A. chartifex* larva; arrow points at the host head. Photos: H. Bahena Basave and G. Pérez-Lachaud.

Gyne parasitism rates. Samples from three nests collected in 2012 containing a total of 1,328 adults (gynes and workers) and 1,329 larvae and pupae were examined (Table 1). All three samples contained parasitized *A. chartifex* gyne larvae and/or free wandering *Nothomicrodon* larvae (Table 1). In general only one *Nothomicrodon* larva develops per host and in only one occasion two parasitoid larvae were observed inside the same host larva (Fig. 4D). A single *Nothomicrodon* puparium was examined; it was more elliptical in body shape than the larva and seemed to have contracted. This puparium had been parasitized and the parasitoid(s) had emerged and gone as revealed by an emergence hole on its surface (see Fig. S4). Rates of *Nothomicrodon* parasitism for the 2012 samples were calculated as the proportion of parasitized gyne larvae with respect to the total number of examined larvae of this caste (in brackets are the corrected values that take into account wandering *Nothomicrodon* larvae and a puparium). Rates were as follows: sample 1: 54.2% (55.6%), sample 2: 0% (30.8%), sample 3: 100%, with an overall proportion of parasitized gyne larvae of 53.9% (57.9%). Larvae from the 2015 samples were not dissected, and estimated gyne parasitism rates were far lower (Table S2), varying from 3.5 to 75.0% with an overall proportion of parasitized gyne larvae of 8.2% (8.6%).

Discussion

In this study we resolve the long-standing enigma of the taxonomic placement of Wheeler's *Nothomicrodon*. Morphological and molecular data reveal that the genus belongs to the Phoridae rather than the Syrphidae where Wheeler²³ had placed it. Furthermore, our data show that these extraordinary myrmecophilous larvae develop as endoparasitoids of *A. chartifex* larvae, and are specific in only developing on the fully-grown gyne larvae.

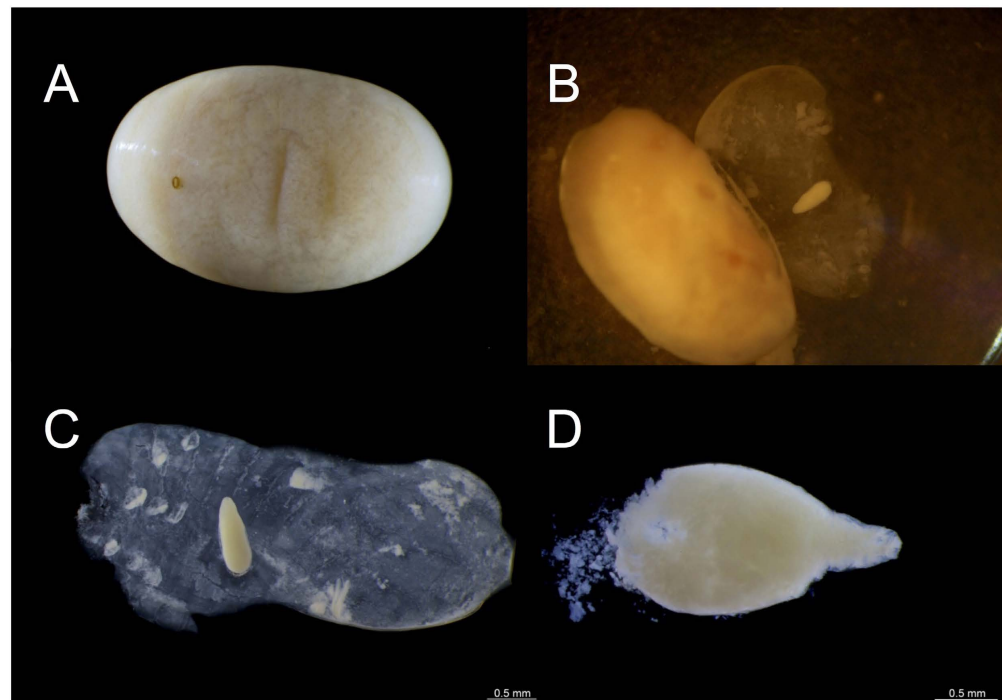


Figure 3. Parasitized *A. chartifex* larva and developmental stages of *Nothomicrodon*. (A) *A. chartifex* larva with sclerotized oviposition scar where the posterior respiratory process of the parasitoid fly larva protrudes. (B) *Nothomicrodon* egg attached to the host cuticle, the host larva has been dissected. (C) Egg. (D) First instar larva dissected from its host. Photos: H. Bahena Basave and G. Pérez-Lachaud.

The larva we studied shares numerous features with that of *N. aztecarum*²³ and both molecular and morphological evidence support placement within the Phoridae. For example, the larval head skeleton is of a platypezoid, not a syrphoid form. Specifically, the apex of the head skeleton is the ventral labial arm which is in the form of a pair of large, sclerotized hooks projecting from the pseudocephalon which are the main food gathering structures in platypezoid larvae²⁸. The DNA sequences identities and the phylogenetic analyses unambiguously show that the larva belongs to the Phoridae. With a rate of similarity of 83 to 88.5%, our sequences, however, are not closely related to any species represented by mtDNA COI sequences in the public sequence databases, and the adult stage remains to be obtained or assessed.

Species boundaries between members of the host genus *Azteca* are not well understood. *Azteca chartifex* belongs to the *A. trigona* group from which the type material of *Nothomicrodon* (*N. aztecarum*) was obtained. It remains possible therefore, that the same species of *Nothomicrodon* is associated with both *A. chartifex* and *A. trigona*.

Phorids are a group of small to minute flies comprising ca. 4,300 recognized species but the more conservative estimates consider that this figure may represent only 10% or less of the total fauna when including undescribed species^{29,31}. They exhibit an array of larval feeding modes, including obligatory and facultative saprophages, predators and parasitoids²⁹. Phorids are parasitic on mollusks and arthropod taxa, such as arachnids, millipedes, and insects. They are well known natural enemies of pest ants³² and adult honey bees³³. Most phorid flies associated with ants live either as nest commensals³⁴, or as parasitoids of foraging workers¹⁹ and occasionally alate females³⁵. Apart from parasitizing ants, phorids also attack other Aculeata, including bees, stingless bees and wasps³. Interestingly, most dipteran parasitoid species attacking social Hymenoptera parasitize the adult stage, although scattered records exist of phorids attacking the larvae of social Hymenoptera (see Tables S3 and S4). About 40% of these cases concern species which develop as ectoparasitoids of formicid and vespidae larvae (see Table S3). Larval endoparasitism by phorids is almost exclusively associated with ants (see Table S4). While only two species of the phorid genus *Aenigmatias* (see Table S3 and references therein) are ectoparasitoids of ant larvae, a growing bulk of records now concerns ant larval endoparasitism by phorids (see Table S4 and references therein). The discovery, in this study, of a *Nothomicrodon* species that is an endoparasite of ant larvae hints that other instances of ant larval parasitism exist in phorids. Our results and literature search reveal that the Phoridae is the only family with parasitoid species that attack both adult ants and their broods with, in the case of *Nothomicrodon*, a specialization for a specific brood caste, i.e. gyne larvae. Several parasitic wasps (Hymenoptera) also attack adult ants or their brood (larvae or pupae), however, this is achieved by distinct wasp families¹².

Several morphological features appear to adapt the *Nothomicrodon* larva to a parasitic feeding mode. The labial hooks facilitate piercing, tearing and loosening fragments of host tissue which are then sucked up by the pump in the head skeleton, and guided towards it by the relatively immobile labrum and at either side of it, the

Nest ident.	Gynes	Gyne pupae	Workers	Worker larvae	Gyne larvae	Total larvae	Parasitized gyne larvae	Gyne parasitism rate (%)	Corrected gyne parasitism rate (%) ^a	Developmental stage of <i>Nothomicrodon</i>	<i>Nothomicrodon</i> L ₃ wandering in the host nest	
Aztc 017	0	0	1295	1170	96	1266	52	54.2	55.6	2 eggs, 13 L ₁₋₂ , 12 early L ₃ , 25 L ₃	3	
Aztc 032	0	0	0	0	9	9	0	0	30.8		3	1 parasitized puparium
Aztc 033	7	44	26	0	10	10	10	100	100	10 L ₃	4	
Total	7	44	1321	1170	115	1285	62	53.9	57.9	62	10	1

Table 1. *Azteca chartifex* material examined for this study, gyne parasitism rate and number and developmental stage of *Nothomicrodon*. ^aCorrected to take into account the free wandering *Nothomicrodon* larvae and a puparium.

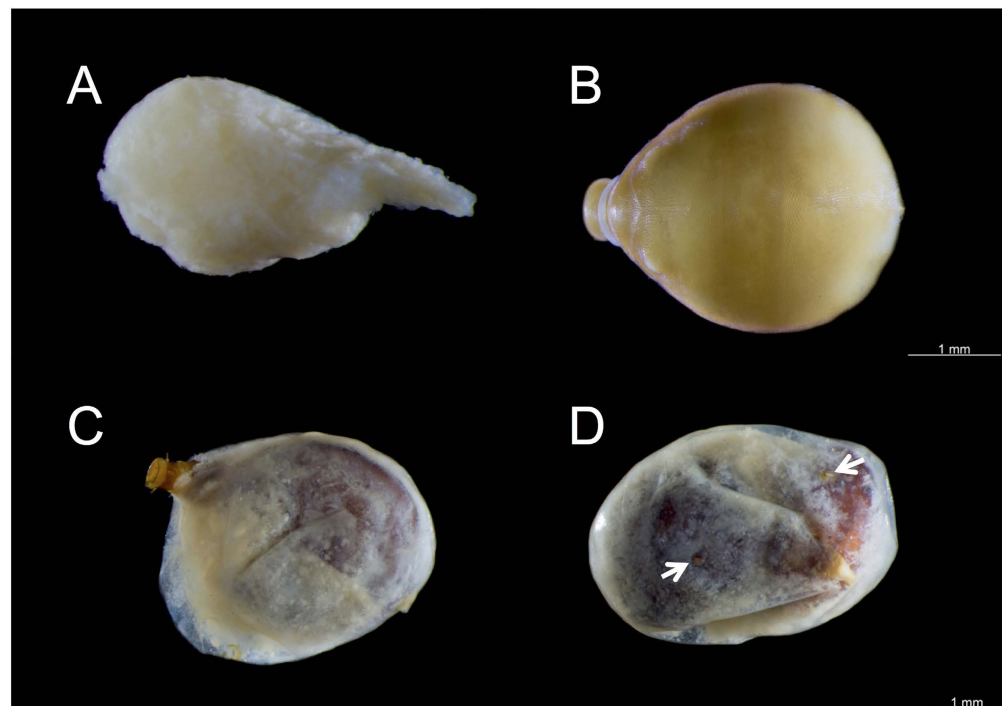


Figure 4. *Nothomicrodon* developmental stages. (A) Second instar larva, dissected from the host larva. (B) Early third instar larva dissected from the host. (C) *Nothomicrodon* third instar larva emerging from the host cuticle remains. (D) Two fully grown *Nothomicrodon* larvae inside a single host larva; arrows point at the scars from which the posterior spiracles protrude. Photos: H. Bahena Basave.

tapered mandibles. The long, parallel dorsal and ventral cornua of the head skeleton suggest that it is covered in short, wide muscles. Such a characteristic delivers a shallow but strong pumping action³⁶, which is typical of many zoophagous larvae³⁷. Perhaps the most distinctive feature of the *Nothomicrodon* larva is its pear shape with an extremely broad, smooth and apparently inflexible abdomen, contrasting with a highly narrowed, tapered, flexible and retractile thorax. Such a shape is also known in larvae of another taxon of cyclorrhaphan endoparasitoids, the Conopidae, whose larvae attack principally the adults of aculeate Hymenoptera³⁸. The pear shape in conopid larvae is adaptive in that the broad abdomen resides in the abdomen of the host while the narrow thorax reaches through the petiole into the thorax to feed on the high density of muscle tissue. The pear shape of the *Nothomicrodon* larva appears to be similarly adapted. Flexibility in the thorax probably facilitates reaching in and around the host body in order to gather food, and might also help egression from the host after completion of larval growth. The *Nothomicrodon* larva might well eat the host remains surrounding its body, as occurs in other parasitoids, such as the braconid wasp *Toxoneuron bicolor* (= *Toxoneuron nigriceps*). In this endoparasitoid, post-egression feeding enhances growth and survival³⁹. However, on the basis of mouthpart structure, it would likely be difficult for the *Nothomicrodon* larva to fragment the host remains, unlike the braconid which has chewing mandibles. In the *Nothomicrodon* larva, the absence of locomotory structures on the ventrum of the abdomen is possibly explained by the position of the larva inside a host where locomotion is not required. The relative size of the anterior respiratory processes is surprising given the position of the thorax inside the host. In

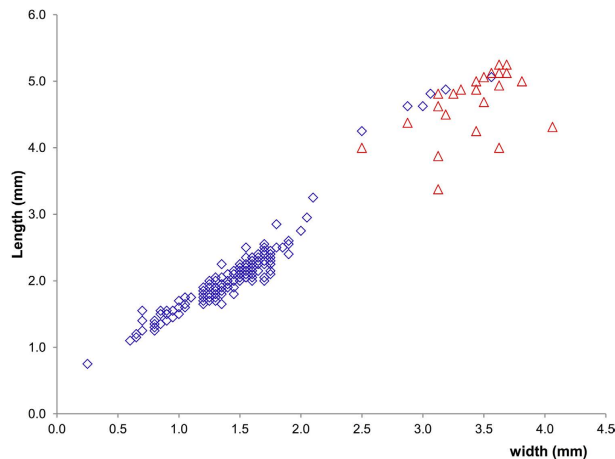


Figure 5. Body length and width of *A. chartifex* larvae. Parasitized larvae in red, unparasitized larvae in blue (n = 212).

contrast, the posterior respiratory process projects only slightly which is probably an adaptive shape in that it is less likely to become caught in host tissue. The flat ventral and slightly convex dorsal surfaces of the larva, together with their hard, leather-like cuticle, and the possibility of retraction of the pseudocephalon and thorax (the most fragile parts of the body) seem to be adaptations to live on the very hard and concave carton walls of the host nest chambers, and might well provide protection from aggressive worker ants.

Whether the *Nothomicrodon* female places its eggs near the host (or host habitat) and the fly larva actively seeks for its host, or lays eggs directly on an *Azteca* ant larva within the host nest remains to be assessed. Whatever, our results show that only the fully-grown gyne larvae of the ant host are targeted, and suggest that host size may be a limiting factor to *Nothomicrodon* larval development. Ant parasitoids impose variable fitness costs on both individuals and colonies^{13,33,40,41}. For high rates of parasitism, parasitoids may significantly reduce resource intake, colony size, and colony fitness. By exclusively parasitizing gyne larvae, *Nothomicrodon* parasitism directly affects the reproductive success of the colony and thereby imposes a high fitness cost to *A. chartifex*. Other parasites and parasitoids impose high reproductive costs on their hosts as for example, in *Nosema* infections of bumble bees⁴². However, fitness costs are not inevitable; not all *A. chartifex* colonies we studied suffered high rates of *Nothomicrodon* parasitism.

Materials and Methods

Insect sampling and preparation. *Azteca chartifex* adults and brood, as well as *Nothomicrodon* larvae, were collected in the state of Bahia in Brazil in 2012 and 2015 (*SI Text: Material and Methods*). *Azteca* workers, larvae and pupae were examined under a stereomicroscope (Nikon SMZ745T) and dissected if parasitized (Table 1). Late instar *Nothomicrodon* larvae found in the nest galleries along with workers and ant brood were collected and examined. Ant larvae collected in 2015 were checked only externally, without dissection, and used essentially for estimating gyne parasitism rates (see Table S4). *Nothomicrodon* larvae and a subsample of *A. chartifex* larvae (including both parasitized and unparasitized larvae) were measured to the nearest 0.1 mm (width × length) using a stereomicroscope provided with an ocular micrometer. A Mixture Discriminant Analysis (MDA) model was fitted to the *A. azteca* data set to test for differences in size between parasitized and unparasitized larvae. MDA is effective for selecting the suitable subclass division of a data set (Gaussian mixture of subclasses)⁴³. The statistical analysis was performed using the MDA package in R⁴⁴. An overall gyne parasitism rate was calculated taking into account all of the potential host larvae examined, with a correction for parasitoid larvae/pupae found freely in the nest chambers. *Nothomicrodon* larvae preserved in alcohol were prepared for description by rehydration and then fixation in Kahle's solution³⁸. To examine the larval head skeleton, the apex of the thorax of a preserved larva was cut off and soaked in hot KOH for about 5 minutes. Excess tissue was removed from the head skeleton and it was washed in acetic acid and stored in glycerol. It was examined using a Wild 5 stereo-microscope in a solid watch glass containing 70% ethanol. The drawing was made with a drawing tube attached to the microscope. Terminology generally follows Rotheray & Gilbert²⁸. Stacked images were obtained using Helicon Focus© (Helicon Soft Ltd). Specimens were also critical point dried and sputter coated before observation with a SM-51 TOPCON Scanning Electron Microscope.

DNA sequencing, identification and clustering. Three second instar larvae of *Nothomicrodon*, obtained by dissection of the hosts, were used for molecular work (labelled Aztc 017B-I, Aztc 017B-II and Aztc 017B-III). DNA was extracted from a small piece of tissue (0.5–1.0 mm sample) of the larvae using the Phire™ Tissue Direct PCR master Mix #F-170S kit (Thermo Scientific Baltics UAB, Vilnius, Lithuania) following the Dilution & Storage protocol with some modifications (*SI Text: Material and Methods*).

The obtained COIa and COIb sequence fragments of our species of *Nothomicrodon*, referred as *incertae sedis* in Table S1, were individually blasted against the BOLD systems v3 (boldsystems.org, accessed 7 March, 2016) and the NCBI GenBank databases, respectively, using BLASTn for the sequence comparisons and identifications.

Sequences produced in this study were deposited in the European Nucleotide Archive (<http://www.ebi.ac.uk/ena>), accession numbers LT592267 (COIa) and LT592268 (COIb).

We additionally used a dataset of COIb sequences retrieved from GenBank with the aim to test the placement of our species of *Nothomicrodon* among samples of the closely related cyclorrhaphan fly families using sequence clustering. The dataset comprised eight COIb sequences of Phoridae species, nine of Syrphidae, five of Platypezidae, four of Pipunculidae, one of Lonchopteridae, and used Sciadoceridae as outgroup (Table S1). The COIb sequence dataset comprised 764 nucleotides and was analyzed using Neighbor-Joining and Maximum Likelihood in software MEGA v.6⁴⁵ using the K2P and General Time Reversible⁴⁶ models of evolution, respectively.

Third instar larvae of our species were further compared to the description and figures of *N. aztecarum* in Wheeler²³ and to the images of the paratype in the Syrphidae Community Website <http://syrphidae.myspecies.info/taxonomy/term/75>. Voucher material of ants and parasitoids was deposited in the following collections: Centro de Pesquisa do Cacau at Ilhéus, Brazil (CPDC collection, CEPEC/CEPLAC) (10 host workers, five third instar *Nothomicrodon* larvae); El Colegio de la Frontera Sur at Chetumal, Mexico (Colección de Formicidae and Colección de Artrópodos) (10 host workers, three third instar *Nothomicrodon* larvae); the National Museums at Edinburgh, Scotland (three third and one second instar *Nothomicrodon* larvae); and the Finnish Museum of Natural History at Helsinki (Finland) (three second instar *Nothomicrodon* larvae, 2 host workers).

References

- Hölldobler, B & Wilson, E. O. *The Ants*. (Springer, Berlin, 1990).
- Kistner, D. H. The social insects' bestiary. *Social Insects* Vol. 3, Hermann, H. R. (ed.), pp. 1–244 (Academic Press, New York, 1982).
- Schmid-Hempel, P. *Parasites in Social Insects*. (Princeton University Press, Princeton, New Jersey, 1998).
- Stoeffler, M., Maier, T. S., Tolash, T. & Steidle, J. L. M. Foreign-language skills in rove-beetles? Evidence for chemical mimicry of ant alarm pheromones in myrmecophilous *Pella* beetles (Coleoptera: Staphilinidae). *J. Chem. Ecol.* **33**, 1382–1392 (2007).
- von Beeren, C., Schulz, S., Hashim, R. & Witte, V. Acquisition of chemical recognition cues facilitates integration into ant societies. *BMC Ecology* **11**, 30 (2011).
- Cushing, P. E. Spider-ant associations: an updated review of myrmecomorphy, myrmecophily, and myrmecophagy in spiders. *Psyche* **2012**, ID 151989 (2012).
- Fiedler, K. The host genera of ant-parasitic Lycaenidae butterflies: a review. *Psyche* **2012**, ID 153975 (2012).
- Powell, S., Del-Claro, K., Feitosa, R. M. & Brandão, C. R. F. Mimicry and eavesdropping enable a new form of social parasitism in ants. *Am. Nat.* **184**, 500–509 (2014).
- Di Giulio, A. *et al.* The pied piper: a parasitic beetle's melodies modulate ant behaviours. *PLoS ONE* **10**, e0130541 (2015).
- Parker, J. Myrmecophily in beetles (Coleoptera): evolutionary patterns and biological mechanisms. *Myrmecol. News* **22**, 65–108 (2016).
- Hughes, D. P., Pierce, N. E. & Boomsma, J. J. Social insect symbionts: evolution in homeostatic fortresses. *Trends Ecol. Evol.* **23**, 672–677 (2008).
- Lachaud, J.-P. & Pérez-Lachaud, G. Diversity of species and behavior of hymenopteran parasitoids of ants: a review. *Psyche* **2012**, ID 134746 (2012).
- Lachaud, J.-P., Klompen, H. & Pérez-Lachaud, G. *Macrodinychus* mites as parasitoids of invasive ants: an overlooked parasitic association. *Sci. Rep.* **6**, 29995 (2016).
- Heraty, J. M. A revision of the genera of Eucharitidae (Hymenoptera: Chalcidoidea) of the world. *Mem. Am. Entomol. Inst.* **68**, 1–368 (2002).
- Thomas, J. A. *et al.* Parasitoid secretions provoke ant warfare. *Nature* **417**, 505–506 (2002).
- Henne, D. C. & Johnson, S. J. Zombie fire ant workers: behavior controlled by decapitating fly parasitoids. *Insect. Soc.* **54**, 150–153 (2007).
- Pérez-Lachaud, G. *et al.* How to escape from the host nest: Imperfect chemical mimicry in eucharitid parasitoids and exploitation of the ants' hygienic behavior. *J. Insect. Physiol.* **75**, 63–72 (2015).
- Lachaud, J.-P. & Pérez-Lachaud, G. Ectaheteromorph ants also host highly diverse parasitic communities: a review of parasitoids of the Neotropical genus *Ectatomma*. *Insect. Soc.* **62**, 121–132 (2015).
- Feener, D. H. Jr. & Brown, B. V. Diptera as parasitoids. *Annu. Rev. Entomol.* **42**, 73–97 (1997).
- Gösswald, K. Pflege des Ameisenparasiten *Tamiclea globula* Meig. (Dipt.) durch den Wirt mit Bemerkungen über den Stoffwechsel in der parasitierten Ameise. *Verh. d. Dtsch. Zool.* **1949**, 256–264 (1950).
- Pérez-Lachaud, G., Jervis, M. A., Reemer, M. & Lachaud, J.-P. An unusual, but not unexpected, evolutionary step taken by syrphid flies: the first record of true primary parasitoidism of ants by Microdontinae. *Biol. J. Linn. Soc.* **111**, 462–472 (2014).
- González, C. T., Wcislo, W. T., Cambra, R., Wheeler, T. A. & Fernández-Marín, H. A new ectoparasitoid species of *Pseudogaurax* Malloch, 1915 (Diptera: Chloropidae), attacking the fungus-growing ant, *Apterostigma dentigerum* Wheeler, 1925 (Hymenoptera: Formicidae). *Ann. Entomol. Soc. Am.* **109**, 629–638 (2016).
- Wheeler, W. M. Two extraordinary larval myrmecophiles from Panama. *Proc. Natl Acad. Sci. USA* **10**, 237–244 (1924).
- Reemer, M. Review and phylogenetic evaluation of associations between Microdontinae (Diptera: Syrphidae) and ants (Hymenoptera: Formicidae). *Psyche* **2013**, ID 538316 (2013).
- Cheng, X.-Y. & Thompson, F. C. A generic conspectus of the Microdontinae (Diptera: Syrphidae) with the description of two new genera from Africa and China. *Zootaxa* **1879**, 21–48 (2008).
- Reemer, M. & Ståhls, G. Generic revision and species classification of the Microdontinae (Diptera, Syrphidae). *ZooKeys* **288**, 1–213 (2013).
- Morehead, S. A., Seger, J., Feener, D. H. Jr. & Brown, B. V. Evidence for a cryptic species complex in the ant parasitoid *Apocephalus paraponerae* (Diptera: Phoridae). *Evol. Ecol. Res.* **3**, 273–284 (2001).
- Rotheray, G. E. & Gilbert, F. Phylogenetic relationships and the larval head of the lower Cyclorrhapha (Diptera). *Zool. J. Linn. Soc.* **153**, 287–323 (2008).
- Disney, R. H. L. *Scuttle Flies: The Phoridae*. (Chapman & Hall, London, UK, 1994).
- Wheeler, G. C. & Wheeler, J. Ant larvae: review and synthesis. *Mem. Entomol. Soc. Wash.* **7**, 1–108 (1976).
- Pape, T., Blagoderov, V. & Mostovski, M. B. Order Diptera Linnaeus, 1758. *Animal biodiversity: An outline of higher-level classification an survey of taxonomic richness*, Zangh, Z.-Q. (ed.), *Zootaxa* **3148**, 222–229 (2011).
- Orr, M. R., Seike, S. H., Benson, W. W. & Gilbert, L. E. Flies suppress fire ants. *Nature* **373**, 292–293 (1995).
- Core, A. *et al.* A new threat to honey bees, the parasitic phorid fly *Apocephalus borealis*. *PLoS ONE* **7**, e29639 (2012).
- Seid, M. A. & Brown, B. V. A new host association of *Commoptera solenopsis* (Diptera: Phoridae) with the ant *Pheidole dentata* (Hymenoptera: Formicidae) and behavioral observations. *Fla Entomol.* **92**, 309–313 (2009).
- Wojcik, D. P., Jouvenaz, D. P. & Lofgren, C. S. First report of a parasitic fly (Diptera: Phoridae) from a red imported fire ant (*Solenopsis invicta*) alate female (Hymenoptera: Formicidae). *Fla Entomol.* **70**, 181–182 (1987).

36. Roberts, M. J. The structure of the mouthparts of some calypterate dipteran larvae in relation to their feeding habits. *Acta Zool.* **52**, 171–188 (1971).
37. Rotheray, G. E. & Wilkinson, G. Trophic structure and function in the larva of predatory muscid flies (Diptera, Muscidae). *Zoomorphology* **134**, 553–563 (2015).
38. Ferrar, P. A guide to the breeding habits and immature stages of Diptera Cyclorrhapha. *Entomograph* **8**, 1–907 (1987).
39. Henderson, R. E., Kuriachan, I. & Vinson, S. B. Postgression feeding enhances growth, survival, and nutrient acquisition in the endoparasitoid *Toxoneuron nigriceps* (Hymenoptera: Braconidae). *J. Insect Sci.* **15**, 51 (2015).
40. Lachaud, J.-P. & Pérez-Lachaud, G. Impact of natural parasitism by two eucharitid wasps on a potential biocontrol agent ant in southeastern Mexico. *Biol. Control.* **48**, 92–99 (2009).
41. Pérez-Lachaud, G., López-Méndez, J. A., Beugnon, G., Winterton, P. & Lachaud, J.-P. High prevalence but relatively low impact of two eucharitid parasitoids attacking the Neotropical ant *Ectatomma tuberculatum* (Olivier). *Biol. Control.* **52**, 131–139 (2010).
42. Otti, O. & Schmid-Hempel, P. A field experiment on the effect of *Nosema bombi* in colonies of the bumblebee *Bombus terrestris*. *Ecol. Entomol.* **33**, 577–582 (2008).
43. Hastie, T., Tibshirani, R. & Friedman, J. *The Elements of Statistical Learning. Data Mining, Inference, and Prediction*, 2nd ed. (Springer, New York, 2009).
44. R Development Core Team 2016. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/> (2016).
45. Tamura, K., Stecher, G., Peterson, D., Filipiński, A. & Kumar, S. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol. Biol. Evol.* **30**, 2725–2729 (2013).
46. Nei, M. & Kumar, S. *Molecular Evolution and Phylogenetics* (Oxford University Press, New York, 2000).

Acknowledgements

We thank Guadalupe Nieto and Humberto Bahena Basave (ECOSUR) for SEM and stereomicroscope imaging, respectively, and Rebeca Gasca for loaning optical equipment. Brian W. Brown provided essential literature on Phorids. J.H.C.D. acknowledges his CNPq research grant and B.J.B.J. acknowledges FAPESPA and FAPESB.

Author Contributions

G.P.-L., B.J.B.J. and J.-P.L. designed the research and analyzed data; B.J.B.J. and J.H.C.D. collected material; G.P.-L. dissected larvae and separated developmental stages; G.S. performed DNA analyses and built the phylogenetic tree; G.R. described the larvae; G.P.-L. and J.-P.L. wrote the first draft of the paper; all authors contributed substantially to the text and revised it.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing Interests: The authors declare no competing financial interests.

How to cite this article: Pérez-Lachaud, G. *et al.* Rediscovery and reclassification of the dipteran taxon *Nothomicrodon* Wheeler, an exclusive endoparasitoid of gyne ant larvae. *Sci. Rep.* **7**, 45530; doi: 10.1038/srep45530 (2017).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2017

Supplementary Information

Rediscovery and reclassification of the dipteran taxon *Nothomicrodon* Wheeler, an exclusive endoparasitoid of gyne ant larvae

Gabriela Pérez-Lachaud¹, Benoit J. B. Jahyny^{2,3}, Gunilla Ståhls⁴, Graham Rotheray⁵, Jacques H. C. Delabie⁶ & Jean-Paul Lachaud^{1,7}

¹ El Colegio de la Frontera Sur, Av. Centenario Km 5.5, Chetumal 77014, Quintana Roo, Mexico

² Universidade Federal do Vale do São Francisco UNIVASF, Colegiado de Ciências Biológicas, Campus Ciências Agrárias - Rodovia BR 407, 12 Lote 543 Petrolina, Pernambuco, Brazil

³ Seção de Entomologia, Comissão Executiva do Plano da Lavoura Cacaueira, Centro de Pesquisa do Cacau (CEPLAC, CEPEC), Cx.P.7, 45600-970, Ilhéus, Bahia, Brazil

⁴ Finnish Museum of Natural History, Entomology Dept., P.O. Box 17, FIN-00014 University of Helsinki, Finland

⁵ National Museums Scotland, West Granton Road, Edinburgh, EH5 1JA, United Kingdom

⁶ Laboratório de Mirmecologia, Convênio CEPLAC/UESC, Cocoa Research Center (CEPEC), 45600-000, Itabuna, Bahia, Brazil

⁷ Centre de Recherches sur la Cognition Animale, Centre de Biologie Intégrative, Université de Toulouse UPS, CNRS-UMR 5169, 118 Route de Narbonne, 31062 Toulouse Cedex 09, France

Content

- SI Text. Materials and Methods.
- Figure S1. SEM micrographs of a *Nothomicrodon* third instar larva.
- Figure S2. Details of a *Nothomicrodon* third instar larva.
- Figure S3. Two-dimensional view of the MDA model fitted to body size measurements.
- Figure S4. *Nothomicrodon* puparium.
- Table S1. List of included taxa and GenBank accession numbers for COIb sequences,
- Table S2. *Azteca chartifex* complementary material.
- Table S3. Phorid ectoparasitoids of social insects larvae,
- Table S4. Phorid endoparasitoids of social insects larvae,
- Table S5. *Azteca chartifex* nests that contained *Nothomicrodon*.

SI Text

Materials and Methods

Ethics Statement

Collection and transport of insect specimens involved in this research were authorized by the Brazilian Chico Mendes Institute for Biodiversity Conservation (SISBIO/ICMBio Permit Number 11304-1 to JHCD). The collection did not involve endangered or protected species. Research and field work comply with the current laws of Brazil.

Azteca chartifex natural history

The dolichoderine genus *Azteca* is an exclusively Neotropical group of arboreal ants, distributed from Mexico to Argentina¹. They are omnivorous, but have a preference for

glucid-rich liquid diets². The biology of the medium sized *A. chartifex* is not well understood yet. This territorial species belongs to the *A. trigona* group, for which various myrmecophilous associates have been reported³. It is common in wet forests of northern South America, from Costa Rica to southern Brazil¹, and dominant in shaded cocoa plantations in Brazil⁴. Colonies are polydomous and occur in clusters of large, conspicuous, pendant carton nests that frequently assume a conical form^{5,6}. The largest unit can extend more than 2 meters in height⁷ and house several thousands of workers. Two groups of workers are clearly distinguished (minors and majors) and physical differences in size are reflected in a clear division of labor, with foragers composed mainly by majors (80%)⁷. Major workers body length is 3.07 ± 0.07 mm⁸. A unique physogastric queen is found in a large chamber near the attachment point on the supporting tree branch; however the queen was observed in only a few nests (B.J.B.J. and J.H.C.D. unpubl. data).

Insect sampling and preparation

Azteca chartifex adults and brood, as well as *Nothomicrodon* larvae, were gathered during a large scale study on ant biodiversity at the Cocoa Research Center (CEPEC-CEPLAC) field station, Ilhéus, Bahia, Brazil. From mid February to October 2012, two *Azteca* nests were collected each month (n = 17). Additionally, at the end of October and early November 2015, nine more nests were harvested for complementary data. Nests were revised in the laboratory in search for myrmecophilous organisms. More specifically, material containing *Nothomicrodon* examined for this study comprised a subset of larvae and adult ants from three nests collected in 2012 and six nests collected in 2015 (see Table S5 for details).

DNA sequencing and identification

The Phire™ Tissue Direct PCR master Mix is designed to perform PCR directly from tissue samples with no prior DNA purification. The tissue sample was placed in an Eppendorf tube in 30 µl of Dilution Buffer, and 0.8 µl of DNA Release Additive was added. The tube was briefly vortexed and centrifuged, incubated at room temperature for about 20 min and then placed in a pre-heated block at 98°C for 2 min, and finally centrifuged at 11 000 rpm for 1 min; 1 µl of supernatant was used in a 20 µl PCR reaction. The cycling conditions were initial denaturation at 98°C for 5 min, 40 cycles of denaturation at 98°C for 5 s, annealing at 49°C for 30 s, extension at 72°C for 20 s, and final extension at 72°C for 1 min. The Folmer fragment or “barcode fragment” of the 5’ region of COI (= COIa) was amplified with forward primer LCO1490 (5’-GCTCAACAAATCATAAAGATATTGG-3’) and reverse primer HCO2198 (5’-TAAACTTCAGGGTGACCAAAAAATCA-3’)⁹, and the COIb fragment was amplified using forward primer C1-J-2183 (5’-CAA CAT TTA TTT TGA TTT TTT GG-3’) (alias JERRY) and the reverse primer TL2-N-3014 (5’-TCC AAT GCA CTA ATC TGC CAT ATT A-3’) (alias PAT)¹⁰. Amplified PCR products were electrophoresed on 1.5% agarose gels and treated with Exo-SapIT (USB Affymetrix, Ohio, USA) prior to sequencing. Both PCR primers were used for sequencing. The Big Dye Terminator Cycle Sequencing Kit (version 3.1) (Applied Biosystems, Foster City, CA, USA) was used on an ABI 3730 (Applied Biosystems, Foster City, CA, USA) genetic analyzer at the Sequencing Service Laboratory of the Finnish Institute for Molecular Medicine (www.fimm.fi). The sequences were edited for base-calling errors and assembled using Sequencher™ (version 5.0) (Gene Codes Corporation, Ann Arbor, MI, USA).

1. Longino, J. T. A taxonomic review of the genus *Azteca* (Hymenoptera: Formicidae) in Costa Rica and a global revision of the *aurita* group. *Zootaxa* **1491**, 1–63 (2007).

2. Harada, A. Y., Benson, W. W. Espécies de *Azteca* (Hymenoptera: Formicidae) especializadas em *Cecropia* spp. (Moraceae): distribuição geográfica e considerações ecológicas. *Rev. Bras. Entomol.* **32**, 423–435 (1988).
3. Wheeler, W. M. Two extraordinary larval myrmecophiles from Panama. *Proc. Natl Acad. Sci. USA* **10**, 237–244 (1924).
4. Majer, J. D., Delabie, J. H. C. & Smith, M. R. B. Arboreal ant community patterns in Brazilian cocoa farms. *Biotropica* **26**, 73–83 (1994).
5. Delabie, J. H. C., Benton, F. P. & Medeiros, M. A. La polydomie chez les Formicidae arboricoles dans les cacaoyères du Brésil: optimisation de l'occupation de l'espace ou stratégie défensive? *Actes Coll. Ins. Soc.* **7**, 173–178 (1991).
6. Delabie, J. H. C. *et al.* Contribution of cocoa plantations to the conservation of native ants (Insecta: Hymenoptera: Formicidae) with a special emphasis on the Atlantic Forest fauna of southern Bahia, Brazil. *Biodivers. Conserv.* **16**, 2359–2384 (2007).
7. Wheeler, D. E. Polymorphism and division of labor in *Azteca chartifex laticeps* (Hymenoptera: Formicidae). *J. Kansas Entomol. Soc.* **59**, 542–548 (1986).
8. Vencl, F. V., Nogueira-de-Sá, F., Allen, B. J., Windsor, D. M. & Futuyma, D. J. Dietary specialization influences the efficacy of larval tortoise beetle shield defenses. *Oecologia* **145**, 404–414 (2005).
9. Folmer, O., Black, M., Hoeh, W., Lutz, R. & Vrijenhoek, R. DNA primers for amplification of mitochondrial cytochrome *c* oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar. Biol. Biotech.* **3**, 294–299 (1994).
10. Simon, C. *et al.* Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Ann. Entomol. Soc. Am.* **87**, 651–701 (1994).

11. Enderlein, G. *Oniscomyia dorni*, eine neue deutsche als Ameisengast lebende flügellose Fliegengattung, sowie über die systematische Stellung der *Thaumatoxena*. *Zool. Jahrb.* **27**, 145–156 (1908).
12. Wasmann, E. Weitere Beiträge zum sozialen Parasitismus und der Sklaverei bei den Ameisen. *Biol. Centralbl.* **28**, 726–731 (1908).
13. Donisthorpe, H. *Aenigmatias blattoides* Meinert, captured in Scotland. *Entomol. Rec.* **25**, 277–278 (1913).
14. Donisthorpe, H. Some notes on the genera *Platyphora*, Verrall, and *Aenigmatias*, Meinert, and a species new to Britain. *Entomol. Rec.* **26**, 276–278 (1914).
15. Donisthorpe, H. St J. K. *The Guests of British Ants. Their habits and life histories.* (G. Routledge & Sons, London, UK, 1927).
16. Falk, S. J. & Chandler, P. J. A review of the scarce and threatened flies of Great Britain. Part 2: Nematocera and Aschiza not dealt with by Falk (1991). *Species Status* **2**, 1–189 (2005).
17. Schmitz, H. Die myrmecophilen Phoriden der Wasmann'schen Sammlung. *Zool. Jahrb.* **37**, 509–566 (1914).
18. Nelson, J. M. Parasites and symbionts of nests of *Polistes* wasps. *Ann. Entomol. Soc. Am.* **61**, 1528–1539 (1968).
19. Richards, O. W. & Richards, M. J. Observations on the social wasps of South America. *Trans. R. Ent. Soc. Lond.* **102**, 1–169 (1951).
20. Jeanne, R. L. Descriptions of the nests of *Pseudochartergus fascatus* and *Stelopolybia testacea* with a note on a parasite of *S. testacea* (Hymenoptera: Vespidae). *Psyche* **77**, 54–69 (1970).
21. Little, M. Social biology of the polistine wasp *Mischocyttarus labiatus*: survival in a Colombian rain forest. *Smith. Contr. Zool.* **327**, 1–27 (1981).

22. Williams, R. N. Insect natural enemies of fire ants in South America with several new records. *Proc. Tall Timbers Conf. Ecol. Anim. Contr. Habit. Manage.* **7**, 123–134 (1980).
23. Williams, R. N. & Whitcomb, W. H. Parasites of fire ants in South America. *Proc. Tall Timbers Conf. Ecol. Anim. Contr. Habit. Manage.* **5**, 49–59 (1974).
24. Packard, A. S., Jr. The parasites of the hobey-bee. *Am. Nat.* **2**, 195–205 (1868).
25. Thompson, F. Some notes on the hunchback fly (*Phora incrassata*, Mg.). *Bee World* **19**, 19–23 (1938).
26. Wheeler, G. C. & Wheeler, J. The ant larvae of the subfamily Formicinae. *Ann. Entomol. Soc. Am.* **46**, 126–171 (1953).
27. Lachaud, J.-P. & Pérez-Lachaud, G. Ectaheteromorph ants also host highly diverse parasitic communities: a review of parasitoids of the Neotropical genus *Ectatomma*. *Insect. Soc.* **62**, 121–132 (2015).
28. Wheeler, G. C. & Wheeler, J. The ant larvae of the subfamily Ponerinae — Part I. *Am. Midl. Nat.* **48**, 111–144 (1952).
29. Wheeler, G. C. & Wheeler, J. The ant larvae of the subfamily Dolichoderinae: Supplement. *Ann. Entomol. Soc. Am.* **59**, 726–732 (1966).
30. Wheeler, G. C. & Wheeler, J. Ant larvae: review and synthesis. *Mem. Ent. Soc. Wash.* **7**, 1–108 (1976).

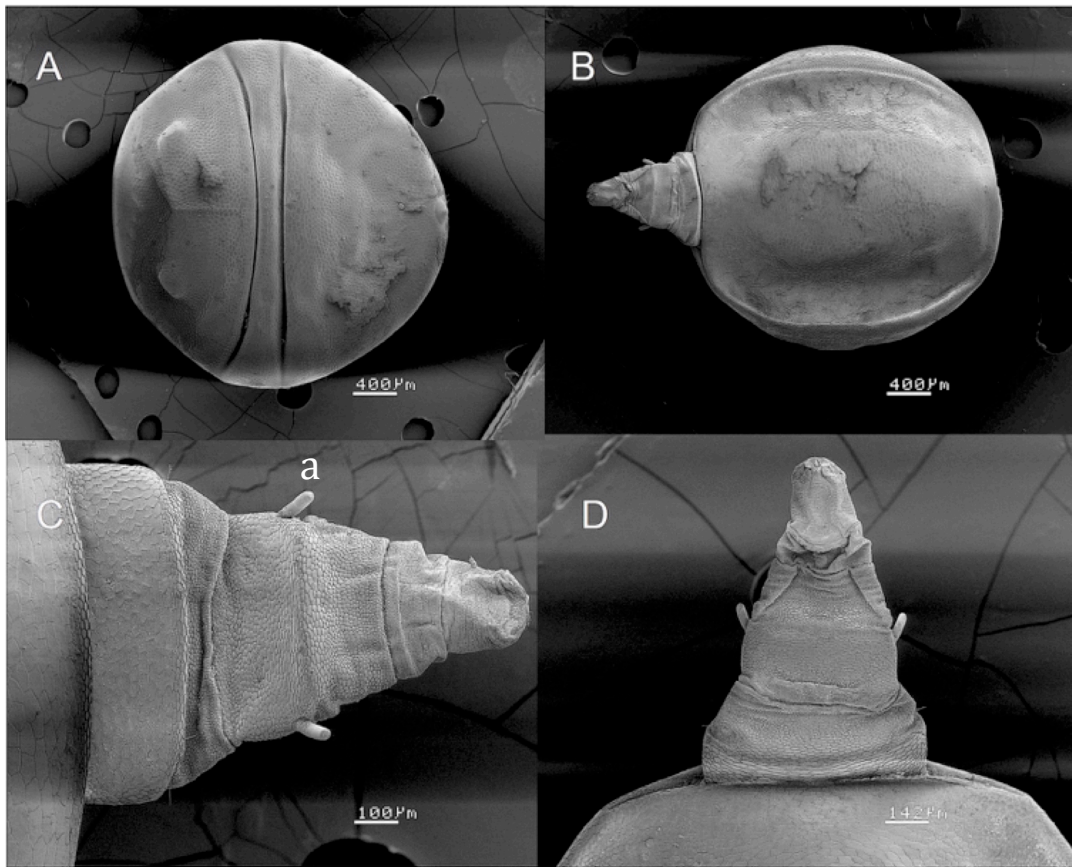


Figure S1. SEM micrographs of a *Nothomicrodon* third instar larva. (A) Whole larva, dorsal view (pseudocephalon and thorax retracted). (B) Whole larva, ventral view. (C) Pseudocephalon and thorax, dorsal view; a: anterior respiratory process. (D) Pseudocephalon and thorax, ventral view. Photos: Guadalupe Nieto.

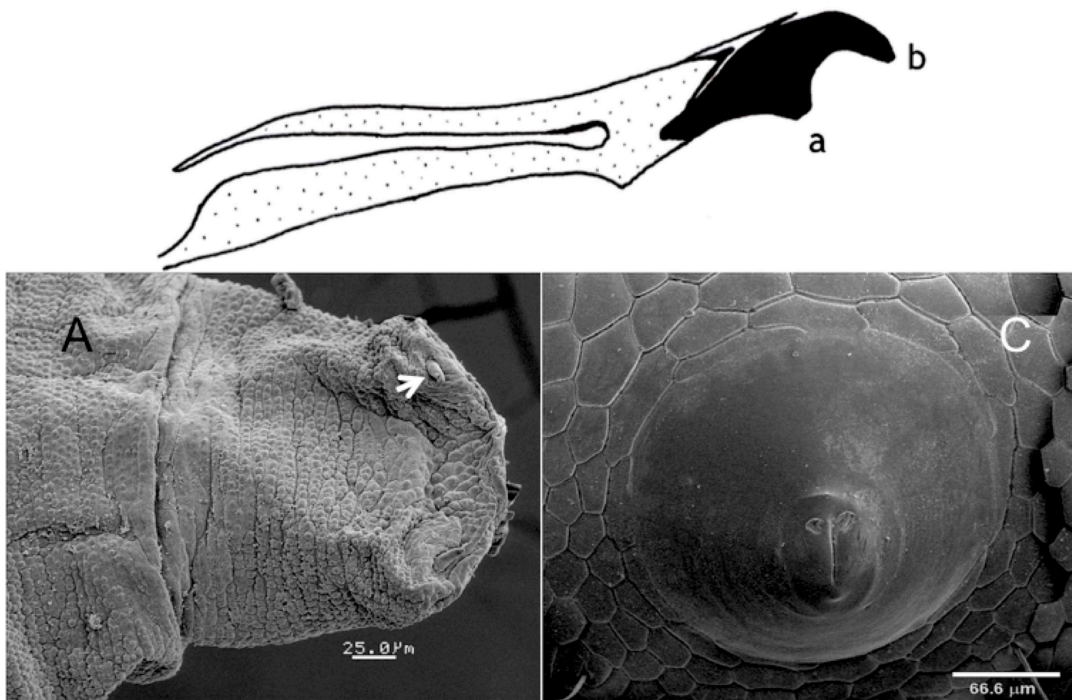


Figure S2. Details of a *Nothomicrodon* third instar larva. (A) Drawing of lateral view of the head skeleton, actual length 0.5 mm. The black hooks are not mandibles, they are the sclerotized apex of the ventral labial arm, a platypozoid feature supporting the molecular work showing that this is a phorid; a: ventral labial arm; b: labial hook. (B) SEM micrograph of pseudocephalon, dorsal view; the arrow points at the left antenna. (C) SEM micrograph of the posterior spiracular system. Photos: Guadalupe Nieto.

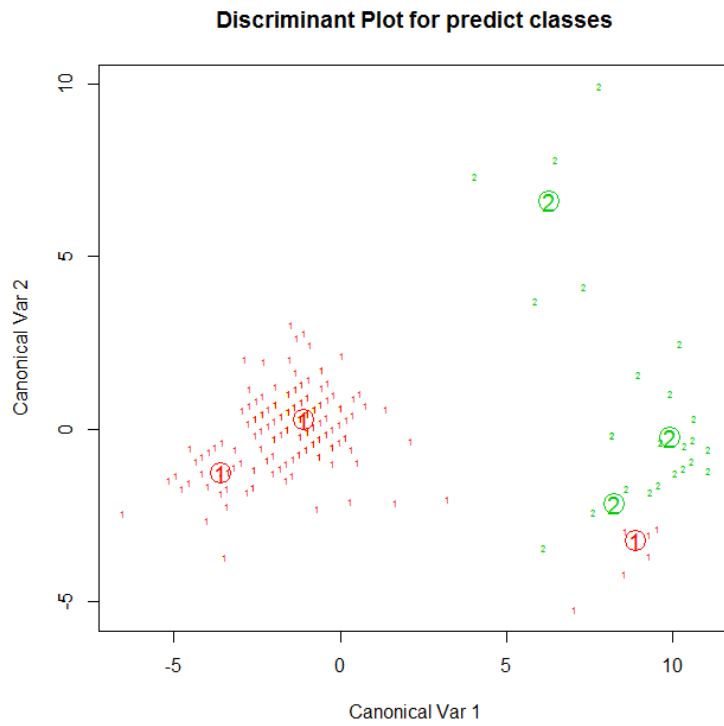


Figure S3. Two-dimensional view of the MDA model fitted to body size measurements. The points are independent sample data projected on to the leading two canonical coordinates. The subclass centers are indicated. Red figures: unparasitized *Azteca chartifex* larvae; green figures: parasitized larvae.

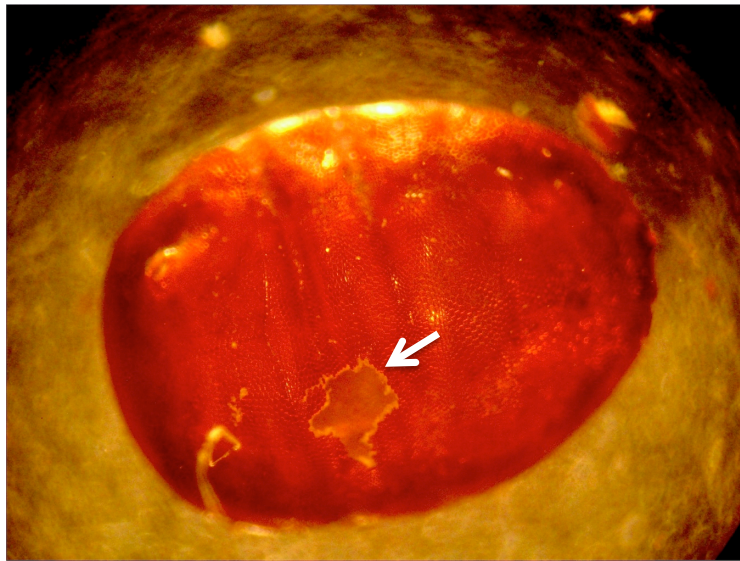


Figure S4. *Nothomicrodon puparium*. This individual has been parasitized; arrow points at the exit hole of the (unknown) parasitoid(s). Photo G. Pérez-Lachaud.

Table S1. List of included taxa and GenBank accession numbers for COIb sequences.

Family	Species	GenBank accession COIb
Lonchopteridae	<i>Lonchoptera tristis</i> Meigen	HM062534
Phoridae	<i>Apocephalus paraponerae</i> Borgmeier	AF217481
	<i>Apodicrania molinai</i> Borgmeier	GU559947
	<i>Diplonevra florea</i> (Fabricius)	HM352612
	<i>Diplonevra setigera</i> Malloch	HM352586
	<i>Dohrniphora anterospinalis</i> Borgmeier	HM352599
	<i>Gymnophora spiracularis</i> Borgmeier	GU559948
	<i>Megaselia</i> sp.	KM224503
	<i>Melaloncha variabilis</i> Brown	EU068515
<i>Incertae sedis</i>	<i>Nothomicrodon</i> sp.	LT592268
Pipunculidae	<i>Chalarus spurius</i> (Fallén)	HS547919
	<i>Eudorylas fuscus</i> (Zetterstedt)	LT797567
	<i>Jassidophaga villosa</i> (von Roser)	AY261685
	<i>Nephrocerus lapponicus</i> Zetterstedt	HF547906
Platypezidae	<i>Agathomyia sexmaculata</i> (von Roser)	KC699450
	<i>Microsania pectipennis</i> (Meigen)	KC699469
	<i>Paraplatypeza bicincta</i> (Szilády)	KC699447
	<i>Platypeza hirticeps</i> Verrall	KC699445
	<i>Polyporivora picta</i> (Meigen)	KM224507
Syrphidae	<i>Cheilosia pedemontana</i> Rondani	AY533336
	<i>Eristalis tenax</i> (Linnaeus)	EU431482
	<i>Metadon captum</i> (Speiser)	LT797565
	<i>Metadon inermis</i> (Loew)	LT797566
	<i>Microdon pictipennis</i> (Macquart)	HF547950
	<i>Merodon albifrons</i> Meigen	DQ386312
	<i>Merodon luteihumerus</i> Marcos García, Vujić & Mengual	LT797564
	<i>Pseudomicrodon</i> sp.	HF547949
<i>Volucella pellucens</i> (Linnaeus)	AY261689	
Sciadoceridae	<i>Sciadocera rufomaculata</i> White	GU559946

Table S2. *Azteca chartifex* complementary material. Collected in 2015, examined for gyne parasitism rate; +: presence; -: absence. As larvae were not dissected, developmental stages of *Nothomicrodon* present in the parasitized larvae could not be secured.

Nest indent.	Gynes	Gyne pupae	Workers	Worker larvae	Gyne larvae	Parasitized gyne larvae	Gyne parasitism rate (%)	Corrected gyne parasitism rate (%) ^a	<i>Nothomicrodon</i> L ₃ wandering in the host nest
2015/1	+	+	+	+	87	32	36.8	40.2	5
2015/2	-	-	-	+	4	3	75.0	75.0	0
2015/3	-	+	+	+	662	42	6.3	6.3	0
2015/4	+	+	+	+	48	33	68.8	70.6	2
2015/7	+	+	+	+	863	30	3.5	3.5	0
2015/8	+	+	+	+	106	5	4.7	4.7	0
Total					1770	145	8.2	8.6	7

^a Corrected to take into account the free wandering *Nothomicrodon* larvae and the presence of a puparium.

Table S3. Phorid ectoparasitoids of social insects larvae.

Ectoparasitoid species	Host species	Host family	Observations	References
<i>Aenigmatias brevifrons</i> (Schmitz) ^a	<i>Formica rufibarbis</i> Fabricius	Formicidae	Exact host identity not fully elucidated	(11–16)
<i>Aenigmatias lubbockii</i> (Verrall) ^b	<i>Formica fusca</i> Linnaeus	Formicidae		(12–14, 16, 17)
	<i>Formica picea</i> Nylander	Formicidae		(14, 16)
<i>Megaselia aletiae</i> (Comstock)	<i>Polistes exclamans</i> Viereck	Vespidae		(18)
<i>Megaselia scalaris</i> (Loew)	<i>Polistes canadensis</i> (Linnaeus)	Vespidae		(19)
	<i>Agelaia testacea</i> (Fabricius) ^c	Vespidae		(20)
<i>Megaselia</i> sp. nr. <i>scalaris</i> (Loew)	<i>Mischocyttarus labiatus</i> (Fabricius)	Vespidae		(21)

^aReferred to as *Platyphora lubbocki* and *P. dorni*. ^bReferred to as *A. blattoides* Meinert and *A. highlandicus* Schmitz. ^cReferred to as *Stelopolybia testacea*.

Table S4. Phorid endoparasitoids of social insects larvae.

Endoparasitoid species	Host species	Host family	Observations	References
<i>Apodicrania termitophila</i> (Borgmeier)	<i>Solenopsis invicta</i> Buren	Formicidae		(22, 23)
<i>Borophaga incrassata</i> (Meigen)	<i>Apis mellifera</i> Linnaeus	Apidae	On fully grown honeybee larvae	(24, 25)
<i>Nothomicrodon</i> sp.	<i>Azteca chartifex</i> Forel	Formicidae		This work
Unidentified phorid species	<i>Formica fusca</i> Linnaeus	Formicidae	6 semipupae each containing a large maggot	(26)
Unidentified phorid species	<i>Ectatomma ruidum</i> (Roger)	Formicidae	2 larvae each containing a fully grown dipteran larva	(27)
Unidentified phorid species	<i>Gnamptogenys tortuolosa</i> (F. Smith)	Formicidae	8/15 larvae containing parasitoid maggots	(28)
Unidentified species (presumably phorid)	<i>Dolichoderus doriae</i> Emery	Formicidae	47/500 larvae contained 1 or 2 dipterous larvae	(29)
Unidentified species (presumably phorid)	<i>Pachycondyla</i> sp.	Formicidae		(30)
Unidentified species (presumably phorid)	<i>Technomyrmex albipes</i> (F. Smith)	Formicidae	75/114 ant larvae contained 1 or 2 dipterous larvae	(29)

Table S5. *Azteca chartifex* nests that contained *Nothomicrodon*. Collected in 2012 and 2015 at CEPEC-CEPLAC (Cocoa Research Center) field station, Ilhéus, state of Bahia, Brazil.

Nest identification	Collecting date	Geographical coordinates	Supporting tree	Height above ground (cm)	Nest length (cm)	Minimum nest width (cm)	Maximum nest width (cm)
Aztc 017	27/04/2012	14°45'52.0"S 39°14'10.1"W	<i>Psidium guajava</i>	350	25; 21 ^a	32; 24 ^a	36; 37 ^a
Aztc 032	21/09/2012	14°46'00.0"S 39°13'28.0"W	<i>Theobroma cacao</i>	60	60	27	35
Aztc 033	11/10/2012	14°46'01.0"S 39°13'28.0"W	<i>T. cacao</i>	150	73	24	45
2015/1	25/10/2015	14°45'22.6"S 39°13'42.9"W	<i>T. cacao</i>	155	40	36	20
2015/2	26/10/2015	14°45'11.3"S 39°13'39.6"W	<i>T. cacao</i>	225	30	26	21
2015/3	26/10/2015	14°45'10.2"S 39°13'35.2"W	<i>T. cacao</i>	200	32	31	14
2015/4	28/10/2015	14°45'18.0"S 39°13'37.5"W	<i>T. cacao</i>	156	30	45	30
2015/7	31/10/2015	14°45'31.6"S 39°13'47.2"W	<i>T. cacao</i>	110	54	35	28
2015/8	01/11/2015	14°45'31.6"S 39°13'47.2"W	<i>T. cacao</i>	150	60	29	24

^aMeasures from two nests: a destroyed nest with few ants on the ground, and a new one constructed in its original place.